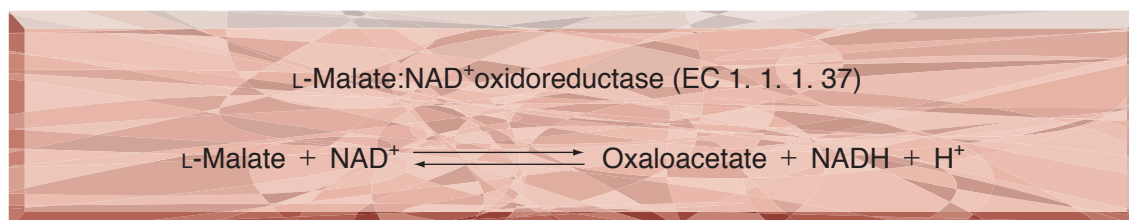


● **TOYOBO ENZYMES** ●  
(Diagnostic Reagent Grade)

# MALATE DEHYDROGENASE

*from Microorganism*



## PREPARATION and SPECIFICATION

Appearance	: Slightly yellowish amorphous powder, lyophilized	
Activity	: Grade II 40U/mg-solid or more	
Contaminants	: Glutamate oxaloacetate transaminase	≤1.0×10 <sup>-3</sup> %
	: Lactate dehydrogenase	≤1.0×10 <sup>-3</sup> %
	: NADH oxidase	≤1.0×10 <sup>-3</sup> %

## PROPERTIES

Stability	: Stable at -20°C for at least one year	(Fig.1)
Molecular weight <sup>1)</sup>	: approx. 140,000	
Isoelectric point <sup>2)</sup>	: pH 4.8±0.1	
Michaelis constants <sup>3)</sup>	: 5.4×10 <sup>-6</sup> M (L-Malate), 5.0×10 <sup>-6</sup> M (Oxaloacetate), 8.1×10 <sup>-6</sup> M (NADH)	
Structure	: 4 subunits per enzyme molecule	
Inhibitors	: Hg <sup>++</sup>	
Optimum pH	: 8.0	(Fig.2)
Optimum temperature	: 70°C	(Fig.3)
pH Stability	: pH 3.0-9.0 (25°C, 20hr)	(Fig.4)
Thermal stability	: below 70°C (pH 7.5, 15min)	(Fig.5)
Effect of various chemicals	: (Table 1)	

## APPLICATIONS

This enzyme is useful for enzymatic determination of L-malate and of glutamate oxaloacetate transaminase (GOT) in clinical analysis.

## ASSAY

### Principle:



The disappearance of NADH is measured at 340nm by spectrophotometry.

### Unit definition:

One unit causes the oxidation of one micromole of NADH per minute under the conditions described below.

### Method:

#### Reagents

- A. K-phosphate buffer, pH 7.5 : 0.1M  
 B. Oxaloacetate solution : 15mM [2.0mg oxaloacetic acid (MW=132.1)/ml of ice-cold K-phosphate buffer (A). This reagent is rather unstable and should be stored in an ice-bath during use] (Should be prepared fresh)  
 C. NADH solution : 6.0mM [4.25mg NADH · Na<sub>2</sub> (ORIENTAL YEAST, MW=709.4)/ml of H<sub>2</sub>O] (Should be prepared fresh)  
 D. Enzyme diluent : 0.1M K-phosphate buffer, pH 7.5 contg. 0.2% BSA

#### Procedure

- Prepare the following reaction mixture in a cuvette (d=1.0cm) and equilibrate at 30°C for about 5 minutes.
 

Concentration in assay mixture	
K-phosphate buffer	97 mM
Oxaloacetate	0.49mM
NADH	0.20mM

- 2.80ml K-phosphate buffer, pH 7.5 (A)  
 0.10ml Oxaloacetate solution (B)  
 0.10ml NADH solution (C)

- Add 0.05ml of the enzyme solution\* and mix by gentle inversion.
- Record the decrease in optical density at 340nm against water for 3 to 4 minutes in a spectrophotometer thermostated at 30°C, and calculate the  $\Delta OD$  per minute from the initial linear portion of the curve ( $\Delta OD$  test).

At the same time, measure the blank rate ( $\Delta OD$  blank) by using the same method as the test expect that the enzyme diluent is added instead of the enzyme solution.

- \* Dissolve the enzyme preparation in ice-cold enzyme diluent (D), dilute to 0.05–0.5U/ml with the same buffer and store on ice.

#### Calculation

Activity can be calculated by using the following formula :

$$\text{Volume activity (U/ml)} = \frac{\Delta OD/\text{min} (\Delta OD \text{ test} - \Delta OD \text{ blank}) \times V_t \times df}{6.22 \times 1.0 \times V_s} = \Delta OD/\text{min} \times 9.807 \times df$$

$$\text{Weight activity (U/mg)} = (\text{U/ml}) \times 1/C$$

V<sub>t</sub> : Total volume (3.05ml)

V<sub>s</sub> : Sample volume (0.05ml)

6.22 : Millimolar extinction coefficient of NADH under the assay condition (cm<sup>2</sup>/micromole)

1.0 : Light path length (cm)

df : Dilution factor

C : Enzyme concentration in dissolution (c mg/ml)

## REFERENCES

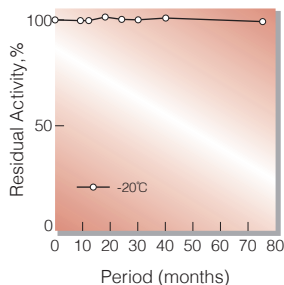
- 1) C.J.R.Thorne and N.O.Kaplan; *J.Biol.Chem.*, **238**, 1861 (1963).
- 2) R.G.Wolfe and J.B.Neilands; *J.Biol.Chem.*, **221**, 61 (1956).
- 3) C.J.R.Thorne; *Biochim, Biophys, Acta.*, **59**, 624 (1962).
- 4) D.J.Blondie et al; *Can.J.Biochem.*, **45**, 641 (1967).

**Table 1. Effect of Various Chemicals on Malate dehydrogenase**

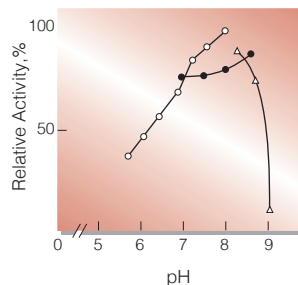
[The enzyme solution dissolved in 0.1M K-phosphate buffer, pH 7.5 contg. 0.2% of BSA (17U/ml) was incubated with each chemical at 25°C for 1hr.]

Chemical	Concn.(mM)	Residual activity(%)	Chemical	Concn.(mM)	Residual activity(%)
None	—	100	2-Mercaptoethanol	2.0	102
Metal salt	2.0		PCMB	0.1	100
MgCl <sub>2</sub>		100	IAA	2.0	99
CaCl <sub>2</sub>		100	Hydroxylamine	2.0	98
Ba(OAc) <sub>2</sub>		101	EDTA	5.0	99
FeCl <sub>3</sub>		102	o-Phenanthroline	2.0	99
CoCl <sub>2</sub>		100	$\alpha, \alpha'$ -Dipyridyl	2.0	100
MnCl <sub>2</sub>		102	Borate	5.0	99
ZnSO <sub>4</sub>		99	NaF	2.0	98
Cd(OAc) <sub>2</sub>		94	NaN <sub>3</sub>	2.0	98
NiCl <sub>2</sub>		100	Triton X-100	0.10%	99
CuSO <sub>4</sub>		99	Brij 35	0.10%	98
Pb(OAc) <sub>2</sub>		99	Tween 20	0.10%	98
AgNO <sub>3</sub>		98	Span 20	0.10%	97
HgCl <sub>2</sub>		0	Na-cholate	0.1%	98
NEM	2.0	100	SDS	0.05%	95
MIA	2.0	99	DAC	0.05%	96

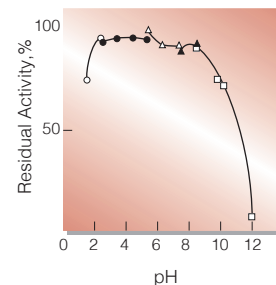
Ac, CH<sub>3</sub>CO; NEM, N-Ethylmaleimide; MIA, Monoiodoacetate; PCMB, p-Chloromercuribenzoate; IAA, Iodoacetamide; EDTA, Ethylenediaminetetraacetate; SDS, Sodium dodecyl sulfate; DAC, Dimethylbenzylalkylammonium chloride.



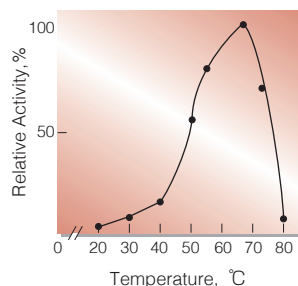
**Fig. 1. Stability (Powder form)**  
[kept under dry conditions]



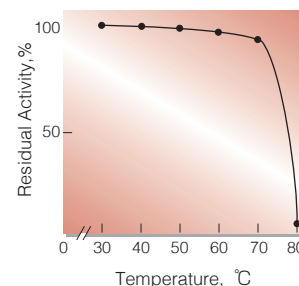
**Fig. 2. PH-Activity**  
[30°C, in 0.1M buffer solution:  
pH5.5-8.0, K-phosphate; pH7.0-8.5, Tris-HCl;  
pH8.0-9.0, Borate]



**Fig. 4. pH-Stability**  
[25°C, 20hr-treatment with 0.1M buffer solution:  
pH2.0-3.5, glycine-HCl; pH3.0-6.0, acetate;  
pH6.0-8.0, K-phosphate; pH8.0-9.0, Tris-HCl;  
pH8.5-12.0, borate]



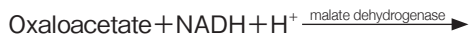
**Fig. 3. Temperature activity**  
[in 0.1mM K-phosphate buffer, pH7.5]



**Fig. 5. Thermal stability**  
[15min-treatment with 0.1m K-phosphate  
buffer, pH7.5  
enzyme concentration: 4.0U/ml]

## 活性測定法 (Japanese)

### 1.原理



NADHの減少を340nmにおける吸光度の変化で測定する。

### 2.定義

下記条件下で1分間に1マイクロモルのNADHを酸化する酵素量を1単位(U)とする。

### 3.試薬

- A. 0.1M K-リン酸緩衝液, pH7.5
- B. 15mMオキサロ酢酸溶液 [2.0mgのオキサロ酢酸 (MW=132.1)を予め氷冷したK-リン酸緩衝液 (A)1.0mlに溶解する] (この試薬はかなり不安定である故,使用時も氷冷保存する)
- C. 6.0mM NADH水溶液 [4.25mgのNADH・Na<sub>2</sub>(オリエンタル酵母製, MW=709.4)を蒸留水1.0mlに溶解する] (用時調製)

酵素溶液：酵素標品を予め氷冷した0.2%牛血清アルブミンを含む50mM K-リン酸緩衝液, pH7.5で溶解し,同緩衝液で0.05~0.5U/mlに希釈して氷冷保存する。

### 4.手順

- ①下記反応混液をキュベット(d=1.0cm)に調製し,30°Cで約5分間予備加温する。
 

2.80ml	K-リン酸緩衝液	(A)
0.10ml	オキサロ酢酸溶液	(B)
0.10ml	NADH水溶液	(C)
- ②酵素溶液0.05mlを添加し,ゆるやかに混和後,水を対照に30°Cに制御された分光光度計で340nmの吸光度変化を3~4分間記録し,その初期直線部分から1分間当りの吸光度変化を求める(ΔOD test)。
- ③盲検は反応混液①に酵素溶液の代わりに酵素希釈液(0.2%牛血清アルブミンを含む50mM K-リン酸緩衝液, pH 7.5)を0.05ml加え,上記同様に操作を行って1分間当りの吸光度変化を求める(ΔOD blank)。

### 5.計算式

$$U/ml = \frac{\Delta OD/min (\Delta OD \text{ test} - \Delta OD \text{ blank}) \times 3.05(ml) \times \text{希釈倍率}}{6.22 \times 1.0 \times 0.05(ml)}$$

$$= \Delta OD/min \times 9.807 \times \text{希釈倍率}$$

$$U/mg = U/ml \times 1/C$$

6.22 : NADHのミリモル分子吸光係数  
(cm<sup>2</sup>/micromole)

1.0 : 光路長(cm)

C : 溶解時の酵素濃度(c mg/ml)